

Stability Study Of Optimized Formulation

Joju Joseph Kattakayam¹, Sangamesh Puranik²

1. Research scholar OPJS University, Churu, Rajasthan, India

2. Research Guide OPJS University, Churu, Rajasthan, India

Submitted: 15-1-2023	Accepted: 10-3-2023

ABSTRACT

Stability Design: In any rational design and evaluation of dosage forms for drug products, the stability of the active component must be a major criterion in determining their acceptance or rejection. Stability can be defined as the ability of a particular formulation, in a specific container, to remain within its physical, chemical, therapeutic and toxicological specifications. Or Stability of a drug can be defined as the time from the date of manufacture and the packaging of the formulation, until its chemical or biological activity is not less than a pre-determined level of labeled potency and its physical characteristics have not changed appreciably or deleteriously. The stability studies were carried as per ICH guidelines. The accelerated study was carried at temperatures of $40^{\circ}C\pm2^{\circ}C/75\%\pm5\%$ RH and real time condition was carried at temperatures of $25^{\circ}C\pm2^{\circ}C/60\%\pm5\%$ RH sample with drawn at respective intervals of 1st month, 2nd month, 3rd month at accelerated condition at real time condition for evaluation parameters such as description, assay, related substances and dissolution profile.

Keywords: Optimized Formulation, T-114, Taste masking, Dissolution Profile, polymethacrylate copolymer

INTRODUCTION:

The present study focus is on the "novel practical approcahes¹⁻⁵ of taste masking of novel antibacterial bitter drug for better patient compliance – some techniques evaluation and characterization".

The drug candidate, Linezolid is an antibacterial drug being used as first line treatment while treating infection. The drug candidate is proven clinically safe and effective via oral route. However, the drug candidate has bitter taste which can't be swollen without masking the bitter taste. The taste needs to be masked in such way that the drug candidate retains its original properties and at the same time taste masked granules of Linezolid will be formulated tablet and also as dry suspension for further evaluation. The objective of this study was to mask the bitter Linezolid using four different techniques with four polymers. Ion exchange resins, Eudragit polymer, betacyclodextrin and glyceryl palmitostearate were used as taste masking agents. Resin drug complexation, Complexation using betacyclodextrin, microsphere using eudragit polymer and melt granulation using glyceryl palmitosterate were the technique followed to assess for the best possible bitter taste masking. The tablets were formulated using direct compression from the drug resin complex and microsphere based granules. The dry suspension was made using betacyclodextrin complex and melt granulated using glyceryl palmitosterate.

Further, The required granules were evaluated for flow properties. The formulated tablets⁶⁻¹⁰ were evaluated for important parameters and also for critical parameters like, dissolution profile, taste evaluation, disintegration, etc. The dry suspension after reconstitution was subjected for critical evaluation like sedimentation volume, viscosity, pH, dissolution profile etc.

The analytical method¹¹⁻¹⁶ was developed and validated meeting the requirements. The quality by design concept was overall evaluated in the present research topic. Also the stability study per ICH guidelines was conducted on the optimized formulations of two tablet two dry suspension formulations.



Table No.1: Stability Design of Taste Masked LinezolidTablets & Dry Powder Suspension:

S No	Test	40°C ± 2°C/75% RH			25°C ± 2°C/60% RH
		1M	2M	3M	3M
1.	Description	~	~	~	~
2.	Assay By HPLC	~	~	~	~
3.	Related substances	~	~	~	~
4.	Dissolution Profile	~	~	~	~

STABILITY STUDY OF OPTIMIZED FORMULATION: ¹⁷⁻¹⁹

Table No. 2. - TEST PARAMETERS TO BEEVALUATED

S. No:	TEST	SPECIFICATION (for information only)
1.	Description	White to off white tables
2.	Assay of by HPLC	Between 90 and 110%
3.	Related substances by HPLC	
	Known Impurities	NMT 0.2% each
	Single maximum unknown Impurity	NMT 0.20%
4.	Total Impurities	NMT 1.0%
5.	Dissolution Profile	NLT 80 Q in 45Minutes

Note: The above test parameters for evaluation is applicable for tablets.

Sl. No.	Test Parameter	Results
1.	Description	Complies
2.	Identification by IR	Complies
3.	Melting point by DSC	180.0°C
4.	Solubility	Selubility Pofile in Common Solvents:
		Solubility (mg/ml)
		Solvent Descriptive term (as per USP)
		Water 2.01 Slightly soluble
		Methanol 20.85 Sparingly soluble
		Dichloromethane 62.65 Soluble
		Dilute Hydrochloric acid 250.10 Freely soluble
5		Quantitative Appears Solubility (mg/m) Descriptive term Solvent Solubility (mg/m) Descriptive term L1-011N HCI 1001 Sparingly soluble 4543PA Accete holfer 2.00 Slightly soluble 6.8435P Toophate huffer 2.86 Slightly soluble 7.5438P Toophate huffer 2.00 Slightly soluble
5.	Loss on Drying	NMT 0.50%
6.	Hygroscopic study	Material was not Hygroscopic
7.	XRD Study	The result conformed to the standard used in the test and conforming to the polymorph

ANALYTICAL METHOD VALIDATION OF ASSAY, RELATED SUBSTANCES AND DISSOLUTION TEST PARAMETERS ¹¹⁻¹⁴

Parameters considered for analytical method validation of Assay method for Linezolid Formulations.

The following parameters were considered for analytical method validation of Assay method in the drug product Linezolid formulations.

System suitability

Specificity

Forced degradation

Precision

System precision

Method precision

Table No. 3: Preformulation Study Results

Indian Journal of Medical and Allied Research



Volume 12, Issue 2, March 2023 pp 8-17. www.ijmar.in ISSN: 2278-0890

Intermediate precision

Stability in analytical solution

Linearity

Accuracy

Range

Robustness

DETAILS OF DRUG SUBSTANCE:

Chemical Structure:

Linezolid:



Figure No.: 1: Structure of Linezolid

Molecular formula and we	eight :	$C_{16}H_{20}FN_3O_4$,	and
337.3461 g/mol			

CAS Number : 165800-03-3

Chemical Name : N-{[(5S)-3-[3-fluoro-4-(morpholin-4-yl) phenyl]-2-oxo-1, 3-oxazolidin-5-yl]methyl}acetamide

REAGENTS, STANDARDS, IMPURITIES AND SAMPLES USED:

Table No.:4 Reagents Details

S.N o.	Reagent
1	Monobasic Sodium Phosphate (NaH ₂ P0 ₄)
2	Acetonitrile
3	Water

Table No.: 5 Details of Standards/Impurities:

S.No.	Standard/Impurity
1	Linezolid working Standard
2	Impurity-B
3	Impurity-C
4	Impurity-D
5	Impurity-I
6	Impurity-II

Table No.: 6: Details of Sample:

S.No.	Sample
1	Linezolid Formulations

Note: All the materials used were within the expiry date and stored at recommended storage conditions.

METHOD DESCRIPTION:

Principle: Reverse phase liquid chromatography with Isocratic elution and UV detector.

Table No.: 7: Chromatographic conditions

Column	:	YMC Hydrosphere C18, 4.6 x 150 mm, 5µ or equivalent
Wavelength	:	254 nm
Injection volume	:	10 µL
Column Temperature	:	25°C
Sample try Temperature	:	25°C
Flow rate	:	1.0 mL/min
Run Time	:	10 minutes
Diluent	:	Mobile phase

Mobile Phase Preparation:

Buffer:

Weigh and transfer 4.7g of monobasic sodium phosphate into 1000 mL of water. Dissolve and mix.



Volume 12, Issue 2, March 2023 pp 8-17. www.ijmar.in ISSN: 2278-0890

Mobile Phase:

Mix 800 mL of buffer and 200 mL of Acetonitrile, Filter and degas by sonication.

Preparation of Standard

Weigh and transfer about 20 mg of Linezolid standard into 50 mL volumetric flask, dilute to volume with diluent and mix well. Further dilute 5.0 mL of this solution to 25 mL with diluent.

Preparation of Sample

Weigh accurately 10 tablets and transfer these tablets into 200 ml amber colour volumetric flask, add 150ml of diluent and sonicate it for 15 minutes; allow it to cool at room temperature. Make up the volume up to the mark with diluent. Dilute 4 ml of this solution to 50 ml with diluent. Filter the sample the sample solution through 0.45 Nylon filter.

Procedure

Inject Diluent (one injection), then inject five replicate injections of standard preparation and check the system suitability parameters.

System suitability: Acceptance criteria

The % RSD for the area of Linezolid replicate injections of standard preparation should be NMT 2.0.

Theoretical plates for Linezolid peak should be NLT 1500.

Tailing for Linezolid peak should be NMT 2.0.

If system suitability parameter passes then inject sample preparation (duplicate injections). Record the area from the chromatograms and calculate the assay.

Calculation and Formulae:

For calculation and formulae, refer section 15.0.

Calculate the % assay of Linezolid by the following formula:

$$\% Assay = \begin{array}{cccc} AT & WS & DT & P & 100 \\ --- & X & ---- & X & ----- & X & ----- \\ AS & DS & V & 100 & LA \end{array}$$

Where,

AT	:	Average area of Linezolid peak from the
		Sample chromatogram.

- AS : Average area of Linezolid peak from the standard chromatogram
- WS : Weight of Linezolid Standard in mg.
- DT : Dilution of sample in mL.

DS	:	Dilution of standard in mL.
V	:	Volume of sample taken (mL)
Р	:	% purity of Linezolid standard
LA	:	Label Amount

SYSTEM SUITABILITY:

To verify the analytical system is working properly and can give accurate and precise results, the system suitability parameters are to be set.

Injected Diluent (one injection) and Standard preparation (5 injections), recorded chromatograms and checked the system suitability parameters.

Table No.: 8: System suitability details

Acceptance criteria	Results
The % RSD for the area of Linezolid replicate injections of standard preparation should be NMT 2.0.	0.1
Theoretical plates for Linezolid peak should be NL T 1500.	10286
Tailing factor for Linezolid peak should be NMT 2.0	1.2

Data interpretation:

From the above results, it can be concluded that the system is suitable for analytical method validation.

SPECIFICITY:

Specificity is the ability of analytical method to assess unequivocally the analyte in the presence of component that may be expected to be present, such as impurities, degradation products and matrix components.

Performed the specificity parameter of the method by injecting Diluent, Standard preparation, Sample preparation> Placebo preparation, known impurities and Sample spiked with impurities into the Chromatographic system and recorded the Retention times.

Acceptance Criteria:

Diluent, placebo and impurities peaks should not interfere with Linezolid peak.

The peaks of Impurities and Linezolid should not interfere with each other.



Volume 12, Issue 2, March 2023 pp 8-17. www.ijmar.in ISSN: 2278-0890

Solutions		Retention time (in min.)
Diluent		-
Placebo preparat	ion	-
Standard prepara	ition	6.193
Sample preparat	ion	6.143
Impurity-I		22.213
Impurity-II		12.933
Impurity-B		4.510
Impurity-C		-
Impurity-D		3.227
Impurity-I		22.073
	Impurity-II	12.860
Sample	Impurity-B	4.503
Sumpre	Impurity-D	3.230
	5-HMF	2.397
	Linezolid	6.153

 Table No.: 9 Retention Time Details

Data interpretation:

From the above results, it can be concluded that there is no interference of diluent, placebo and impurities peaks with Linezolid peak.

SPECIFICITY BY DEGRADATION STUDIES:

Specificity by forced degradation:

Forced degradation of Linezolid injection has been carried out, to confirm that during stability study or throughout the shelf life, any degradation if found should not interfere with the Linezolid peak. In addition the forced degradation study will help to identify the type of degradation pathway (whether oxidative, alkali hydrolysis, acid hydrolysis, water hydrolysis, photolytic and dry heat) for each of the degradants.

Preparation of Sample:

Sample as such: Taken 2 mL sample solution into 50 mL volumetric flask, dissolved, and diluted to volume with diluent.

Placebo as such: Taken 2 mL Placebo solution into 50 mL volumetric flask, dissolved, and diluted to volume with diluent.

Neutral Stressed sample:

Taken 2 mL Sample solution into 50 mL volumetric flask, added 2 mL water to it and kept it in water bath at 80°C for 4 hours. Allowed it to attain room temperature, then diluted to the volume with diluent.

Acid Stressed sample:

Taken 2 mL Sample solution into 50 mL volumetric flask, added 2 mL 0.1 N HCI to it and kept it in water bath at 80°C for 4 hours. Allowed it to attain room temperature. Neutralized with 2 mL of 0.1N NaOH, then diluted to the volume with diluent.

Repeated the same with IN HCl.

Alkali Stressed sample:

Taken 2 mL Sample solution into 50 mL volumetric flask, added 2 mL O.IN NaOH to it and kept it in water bath at 80° C for 4 hours. Allowed it to attain room temperature. Neutralized with 2 mL of 0.1 N HCL, then diluted to the volume with diluent.

Repeated the same with IN NaOH.

Peroxide Stressed sample:

Taken 2 mL Sample solution into 50 mL volumetric flask, added 2 mL 3% Peroxide to it and keep it in room temperature for 7 days. After 7 days, diluted to volume with diluent and mixed well.

Sunlight exposed sample:

Taken 2 mL Sample solution into 50 mL volumetric flask and exposed to Sunlight for 8 hours and diluted the volume with diluent.

UV light exposed sample:

Taken 2 mL Sample solution into 50 mL volumetric flask and exposed to an illumination of 1.2 million Lux hours of cool fluorescent light and an integrated near UV energy exposure of 200-Watt hours/m², simultaneously in photo stability chamber maintained at 25°C.Then dilute to volume with diluent.

Thermal Stressed (Dry heat) sample:

Taken 2 mL Sample solution into 50 mL volumetric flask and exposed to hot air oven at 80°C for 8 hours, cooled to room temperature then diluted the volume with diluent.

Note: The same treatment should be made for placebo.



Acceptance criteria:

Peak of Linezolid should be pure.

All known and unknown impurity/Degradation products if any should be well separated from the Linezolid peak.

Peak purity factor for Linezolid peak should be NLT 0.990.

Table No.: 10. Forced degradation study compilation

Stressed Condition	Linezolid (in Assay%)	Linezolid (% of Degradation)
Sample As such	101.9	-
Alkali	76.4	25.0
Acid	72.1	29.2
Peroxide	91.9	9.7
Neutral	98.9	2.9
Sun light	102.3	-
UV-light	101.5	0.4
Thermal	101.6	0.3

Data Interpretation:

The Sample is found to be degrading in acid, alkali, peroxide and neutral stressed conditions but slightly degraded in UV, sunlight and thermal stressed condition. However, unknown impurities are well separated from Linezolid peak and impurities. The Linezolid peaks are pure. Hence, the Assay method is considered specific & stability indicating.

PRECISION:

The precision of an analytical method is the degree of agreement among individual test results when the method is applied repeatedly to multiple sampling of homogeneous sample. The precision of analytical method is usually expressed as the standard deviation or relative standard deviation (Coefficient of variation) of series of measurements.

SYSTEM PRECISION:

The system precision is checked by using standard chemical substance to ensure that the analytical system is working properly. The retention time and the area response of six determinations should be measured and calculated relative standard deviation. Injected diluent one injection and standard preparation six injections into the chromatograph. Recorded and calculated relative standard deviation.

Acceptance criteria:

The %RSD of the Retention time for the Linezolid peak obtained from 6 injections of standard preparation should be NMT 1.0

The %RSD of the Area response for the Linezolid peak obtained from 6 injections of standard preparation should be NMT 2.0

METHOD PRECISION:

In method precision, a homogeneous sample of a single batch should be analyzed six times. This indicates whether a method is giving consisting results of a single batch.

Analyzed the samples of Linezolid formulations six times of a same batch as per analytical procedure. Calculated the % Assay of Linezolid.

Acceptance Criteria:

The% RSD of the calculated Assay results for 6 determinations should be NMT 2.0

Table No.:11. Percentage assay results of Linezolid in assay test parameter validations

Sample	% Assay of Linezolid
1	101.1
2	101.5
3	101.5
4	101.5
5	101.3
6	101.2
Mean	101.4
% RSD	0.2

Data Interpretation:

From the above results, it can be concluded that the method is precise.



Volume 12, Issue 2, March 2023 pp 8-17. www.ijmar.in ISSN: 2278-0890

STABILTY IN ANALYTICAL. SOLUTION:

Evaluate the stability in analytical solution by injecting the standard preparation and sample preparation at regular interval.

Acceptance criteria:

The % difference of Area Response for the peaks in Standard preparation and Sample preparation should be within $\pm 2.0\%$ from initial area after specified period.

Standard and samp1e preparation stability at 25°C:

Table No.: 12: Solution Stability Details

Linezolid					
Standard preparation		Sample preparation			
Time (in hours)	Area response	% Difference	Time (in hours)	Area response	% Difference
Initial	2648454	-	Initial	2632827	
3	2630435	-0.7	2	2621526	
6	2631686	-0.6	5	2623360	
8	2634353	-0.5	7	2625299	
10	2633493	-0.6	10	2624169	
13	2635801	-0.5	12	2628053	
15	2634647	-0.5	14	2629503	
17	2634510	-0.5	17	2631194	
20	2628064	-0.8	19	2623687	
22	2633645	-0.6	21	2630006	
24	2634786	-0.5	24		
27	2634814	-0.5	26		
29	2634183	-0.5	28		
32	2635942	-0.5	31		

Data Interpretation:

From the above results, it can be concluded that the Standard preparation is stable for 32 Hours at 25°C (%Difference is - 0.5%), Sample preparation is stable up to 31 Hours at 25°C (%Difference is -0.2).

5.8.2.7 LINEARITY:

The linearity of an analytical method is its ability to elicit test results that are directly or by a well-defined mathematical transformation, proportional to the concentration of analyte in samples within a given range.

Performed the linearity with Linezolid standard in the range of 50 to 150% of specification limit.

Recorded the area response for each level and calculated slope, intercept & correlation coefficient. Tested the intercept for statistical equivalence to zero.

Plotted a graph of Linezolid concentration (ppm) on X-axis and Area response on Y -axis.

Table No.:13. Linearity Details

Level	Concentration	Area Posponso	
	ш ррш	Response	
0	0.0000	0	
1	39.2117	1243904	
2	47.0540	1525990	
3	56.8569	1768033	
4	64.6992	2034005	
5	72.5416	2271984	
6	80.3839	2511890	
7	88.2262	2757541	
8	96.0685	3049760	
9	103.9109	3313608	
10	119.5955	3824363	
Correla	ation coefficient	1.000	
Regression coefficient		0.999	
Slope		31786.712	
Intercept		- 11686.235	
% Intercept		-0.5	



Indian Journal of Medical and Allied Research

Volume 12, Issue 2, March 2023 pp 8-17. www.ijmar.in ISSN: 2278-0890



Fig No.: 2. Linearity graphical representation

ACCURACY:

The accuracy of an analytical method is the closeness of test results obtained by that method to the true value (standard value).

Spiked known quantity of Linezolid standard at 50%, 80%, 100%, 120% and 150% of Assay specification limits into the placebo.

Performed precision at the lowest and the highest levels and for the other levels prepared in triplicate and injected in duplicate for all levels. Calculated the % recovery from the results of Accuracy.

% Leve l (abo ut)	Sa mp le	Mean Area Response	*m g/m L Add ed	*mg/m L Recov ered	% Recov ery	Mean % Recov ery	%R SD
	1	1240140		0.0384	98.0		
	2	1240095		0.0384	98.0		
50	3	1243129	0.03	0.0385	98.2	98.0	0.1
50	4	1239626	92	0.0384	98.0	90.0	0.1
	5	1239616		0.0384	98.0		
	6	1240746		0.0384	98.0		
	1	2505096	0.07	0.0776	99.0		
100	2	2501077	84	0.0774	98.7	98.9	0.2
	3	2502763	01	0.0775	98.9		
	1	3806360		0.1178	100.2		
	2	3815696		0.1181	100.4		
150	3	3821258	0.11	0.1183	100.6	100.4	0.1
100	4	3811428	76	0.1180	100.3	100.1	0.1
	5	3817426		0.1182	100.5		
	6	3811600		0.1180	100.3		

 Table No.: 14: Recovery of Linezolid:

Data Interpretation:

From the above results, it can be concluded that the recovery is well within the limit. Hence the Method is accurate.

RANGE:

The range of analytical method is the interval between the upper and lower levels of analyte that has been demonstrated to be determined with a suitable accuracy and linearity. Derived the specified range from the Linearity and Accuracy studies.

Acceptance criteria:

The %RSD obtained for all accuracy level determinations should be NMT 2.0. The Correlation coefficient should be NL T 0.998 for Linearity and Accuracy level determinations.

RESULTS:

Table No.: 15: Linearity Range of Linezolid:

% Level	Concentration in ppm	Mean Area response
50	39.2117	1243904
100	80.3839	2511890
150	119.5955	3824363
	Correlation coefficient	1.000



Fig No.3: Linearity range graphical representation



Table No.: 16: Accuracy Range of Linezolid:

Accuracy Range in Linezolid			
% Level	Mcg/mL added	Mcg/mL recovered	
50	0.0392	0.0384	
100	0.0784	0.0775	
150	0.1176	0.1181	
	Correlation coefficient	1.000	
	%RSD	1.0	



Fig No.4: Accuracy range graphical representation

Data Interpretation:

From the above results, it can be concluded that the Range of the method is from 50% to 150% of target concentration for Linezolid.

ROBUSTNESS:

The robustness of an analytical method is a measure of its capacity to remain unaffected by small but deliberate variations in method parameters and provides an indication of its reliability during normal usage.

Robustness parameters:

Change in column oven temperature $\pm 5^{\circ}$ C.

Change in flow rate ± 0.2 mL/min.

Change the organic phase ratio $\pm~5.0~\%$

Acceptance criteria:

The System suitability parameters should pass for all the conditions.

System suitability Parameter		%RSD for Area	Theoretical plates	Tailing factor
Limit		NMT 2.0	NLT 1500	NMT 2.0
Original cond	Original conditions		10286	1.2
Flow rate	1.2 mL/min	0.0	9213	1.2
	0.8 mL/min	0.1	11423	1.2
Column	30°C	0.1	10795	1.2
Temperature	20°C	0.1	9680	1.2
Organic	+5%	0.2	9938	1.2
Phase	-5%	0.1	10065	1.2

Table No.17: Robustness parameter Details

Data Interpretation: From the above results, it can be concluded that the Method is robust.

CONCLUSION:

The proposed HPLC method for estimation of Assay of Linezolid in the Linezolid formulation is validated. The method is found to be specific. The method is also stability indicating as evidenced by forced degradation studies. The method is found to be linear in the specified range for Linezolid. Accuracy of this method is established for Linezolid. The method is found to be precise and robust. A system suitability test is established and related parameters are recorded. Hence this method stands validated and can be used for routine and stability analysis.

REFERENCES:

1. Jeong SH, Haddish NB, Haghighi K and Park K: Drug release properties of polymer coated ion-exchange resin complexes: experimental and theoretical evaluation. J. Pharm. Sci. 2007; 96: 618–632.

2. Reacher, M. H., Shah, A., Livermore, D. M., Wale, M. C., Graham, C., Johnson, A. P. et al. (2000). Bacteraemia and antibiotic resistance of its pathogens reported in England and Wales between 1990 and 1998: trend analysis. British Medical Journal 320, 213–6.

3. Woodford, N. & Livermore, D. M. (2001). Can we beat MRSA now we know its genome sequence? Lancet Infectious Diseases 1,9–10.

4. Johnson, A. P., Warner, M., Broughton, K., James, D., Efsratiou, A., George, R. C. et al. (2001). Antibiotic susceptibility of streptococci and related genera causing endocarditis: analysis of UK reference laboratory referrals,



January 1996 to March 2000. British Medical Journal322, 395-6.

5. European Antimicrobial Resistance Surveillance System (EARSS) Web site. (2002). Database published by RIVM/EARSS. [Online.] http://www.earss.rivm.nl (30 January 2003, date last accessed).

6. Slee, A. M., Wuonola, M. A., McRipley, R. J., Zajac, I., Zawada, M. J., Bartholomew, P. et al. (1987). Oxazolidinones, a new class of synthetic antibacterial agents: in vitro and in vivo activities of DuP105 and DuP 721. Antimicrobial Agents and Chemotherapy 31, 1791–7.

7. Ford, C. W., Hamel, J. C., Stapert, D., Moerman, J. K., Hutchinson, D. K., Barbachyn, M. R. et al. (1997). Oxazolidinones: new antibacterial agents. Trends in Microbiology5, 196–200.

8. Zurenko, G. E., Yagi, B. H., Schaadt, R. D., Allison, J. W., Kilburn, J. O., Glickman, S. E. et al. (1996). In vitro activities of U-100592 and U-100766, novel oxazolidinone antibacterial agents. Antimicrobial Agents and Chemotherapy 40, 839–45

9. Jones, R. N., Anderegg, T. R. & Deshpande, L. M. (2002). AZD2563, a new oxazolidinone: bactericidal activity and synergy studies combined with gentamicin or vancomycin against staphylococci and streptococcal strains. Diagnostic Microbiology and Infectious Disease 43, 87-90.

10. Cooper J, Gunn C. Powder flow and compaction. In: Carter SJ, eds. Tutorial Pharmacy. New Delhi, India: CBS Publishers and Distributors; 1986:211-233.

11. Shah D, Shah Y, Rampradhan M. Development and evaluation of con- trolled release diltiazem hydrochloride microparticles using cross-linked poly(vinyl alcohol). Drug Dev Ind Pharm. 1997;23(6):567-574.

12. Aulton ME, Wells TI. Pharmaceutics: The Science of Dosage Form Design. London, England: Churchill Livingstone; 1988.

13. Martin A. Micromeritics. In: Martin A, ed. Physical Pharmacy. Balti- more, MD: Lippincott Williams & Wilkins; 2001:423-454.

14. Blase, C.M., Shah, M.N., Taste masked pharmaceutical suspensions for pharmaceutical actives. Eur. Pat. Appl. EP0556057, 18 August 1993.

15. Rathbone, M. J., Hadgraft, J. Drugs & pharmaceutical Sciences in drug delivery technology. Marcel Dekker, New York, 126, 2003 p191-202.

16. Agarwal, R., Mittal, R., Singh, A., Studies of Ion-Exchange Resin Complex of Chloroquine Phosphate. Drug Dev. Ind. Pharm. 6, 2000 p 773-776.

17. S.Mahesh, Formulation & Evaluation of Isopropyl Antipyrine oral suspension Indian j. pharme. Sci.1998; 60(2) 99-94.

18. Geetha Rao the taste masked oral suspension of Quinine Sulphate By Complexation; Indian j.pharme. Sci May – June 2004 329-331.

19. ICH: Q8, Q9 & Q10.